

Mechanical Properties of Microcapsules Used in a Self-Healing Polymer

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Abstract

The elastic modulus and failure behavior of poly(urea-formaldehyde) shelled microcapsules were determined through single-capsule compression tests. Capsules were tested both dry and immersed in a fluid isotonic with the encapsulant. A shell-theory model for a fluid-filled microcapsule was utilized to extract the modulus of the shell wall material from individual capsule tests. The testing of capsules immersed in a fluid had little influence on mechanical behavior in the elastic regime. The average capsule shell wall modulus was determined to be 3.7 GPa. Capsule diameter was found to have a significant effect on burst strength, with smaller capsules sustaining higher stresses before burst.

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I. INTRODUCTION

Microcapsules containing liquid healing agent are a critical component of self-healing polymers [1, 2]. Healing is accomplished by incorporating the microencapsulated healing agent and a catalyst within an epoxy matrix. An approaching crack ruptures embedded microcapsules, releasing healing agent into the crack plane through capillary action. Polymerization of the healing agent is initiated by contact with the embedded chemical catalyst, bonding the crack faces. The rupture of microcapsules is the mechanical trigger to the healing process and without it, no healing occurs. This system has proven to be highly effective at healing cracks in both quasi-static [1] and fatigue [3–6] loading.

An optimal combination of microcapsule and matrix properties is necessary to ensure mechanical triggering when the material is damaged; if the shell wall is too thick the microcapsule will not rupture readily, preventing the release of healing agent. On the other hand, if the shell wall is too thin, the capsules not only are fragile, but also allow diffusion of the healing agent into the matrix. Other key parameters for efficient healing agent delivery are the elastic stiffness, the fill content, and the burst strength of the capsules.

The complex three-dimensional crack–microcapsule interaction which occurs in a self-healing composite has been studied using the Eshelby–Mura equivalent inclusion method [1]. Model predictions reveal that the capsule-to-matrix stiffness ratio influences the crack propagation path in close proximity to the capsule. A capsule with a higher elastic modulus than the surrounding matrix creates a stress field that tends to deflect the crack away from the microcapsule. Conversely, a more compliant shell wall material produces a stress field that attracts the crack toward the microcapsule, facilitating capsule rupture.

In addition to providing storage of the healing monomer, Brown et al. [7] demonstrated that microcapsules toughen the polymer matrix by as much as 127% over the neat matrix (with no capsules) value. Previous studies of microcapsule toughening included only the effect of average capsule diameter and volume fraction. The additional influences of shell wall thickness, capsule processing, and fill content on toughening in a polymer matrix were examined in more recent work [8]. While all three of these parameters significantly impacted the efficiency of microcapsule toughening, it was difficult to elucidate the relationship between physical properties of the microcapsule and the fracture performance of the polymer composite. The goal of the present work is to characterize the mechanical properties of the microcapsule system currently used in self-healing composites.

Several methods for characterizing capsules have appeared in the literature, most of which were developed for biological cells. Mitchison and Swan [9] introduced a micropipette aspiration test for probing the mechanical response biological cells. A micropipette was placed in contact with the cell and a vacuum was applied, drawing the cell wall into the micropipette. The cell wall deflection was measured and related to the mechanical stiffness of the wall material. The micropipette aspiration technique has also been applied to determine microcapsule shell wall elastic modulus [10]. Cole used a simple compression test to characterize the stiffness of sea urchin eggs [11]. A single egg was placed between two platens and then compressed. In addition to characterizing biological cells, this experimental technique has been applied to microcapsule systems [12–14]. In the current work, the single-capsule compression experiment is adopted for the characterization of a range of microcapsules and the resulting load–displacement data compared with a shell theory model to determine the elastic properties of the shell wall.

II. EXPERIMENTAL PROCEDURE

A. Capsule Manufacture

The capsules examined in this study had a poly(urea-formaldehyde)(UF) shell wall and were filled with dicyclopentadiene (DCPD) liquid monomer. They were identical to those used for the self-healing system in [1] and were manufactured by an *in-situ* microencapsulation method. *In-situ* microencapsulation proceeds in two concurrent steps. First a UF prepolymer is formed in an aqueous bath containing an emulsified, water-immiscible encapsulent fluid [15]. The UF polymerizes around an individual DCPD droplet in the emulsion, forming the shell wall of an individual microcapsule. Capsule diameter is determined by the droplet size of the emulsion. A schematic of the manufacture procedure for UF capsules is presented in Fig. 1. The average diameters for each capsule group tested were $213\pm 12\ \mu\text{m}$, $187\pm 15\ \mu\text{m}$, and $65\pm 7\ \mu\text{m}$.

A representative scanning electron microscopy (SEM) micrograph of the surface morphology of a UF capsule is shown in Fig. 2. In addition to allowing investigation of surface morphology, SEM allowed measurement of the shell wall thickness of an individual capsule. These microcapsules possessed a highly uniform shell wall thickness, $175\pm 33\ \text{nm}$, independent of capsule diameter.

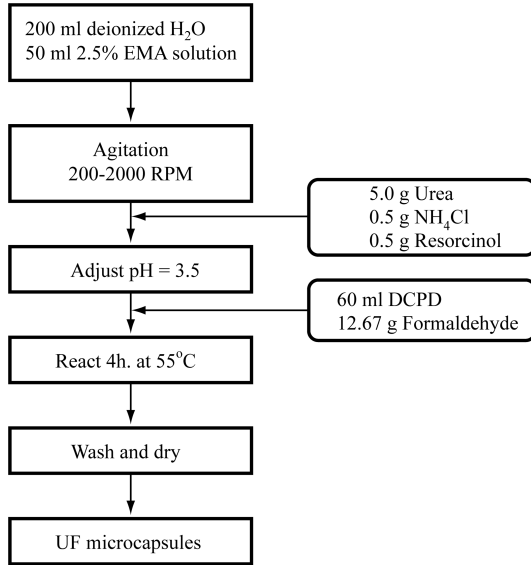


FIG. 1: Encapsulation procedure for UF capsules [15]

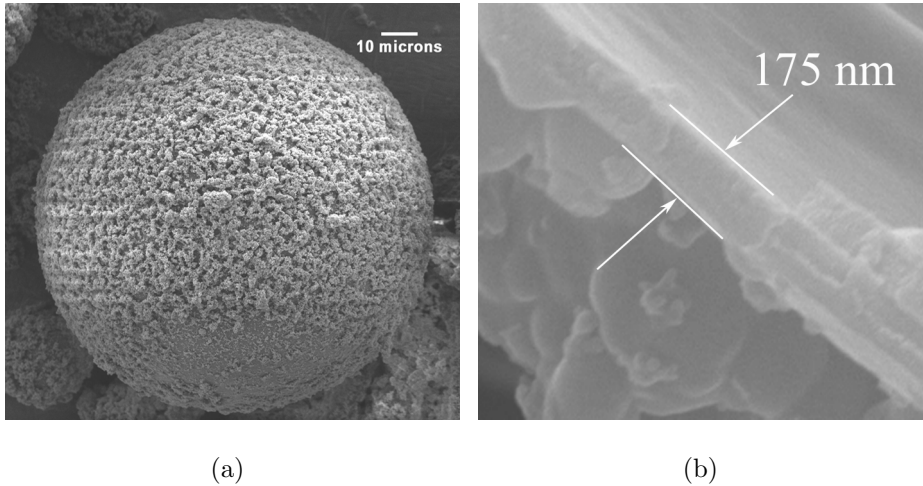


FIG. 2: Electron micrographs of a UF DCPD filled microcapsule: (a) surface morphology; (b) cross-section of the shell wall.

B. Experimental Setup

The capsule compression apparatus, shown in Fig. 3, was adapted from the one described by Liu and coworkers [12]. Displacement was applied at a rate of $5 \mu\text{m/s}$ for the $187 \mu\text{m}$ and $213 \mu\text{m}$ capsule size ranges and $2.5 \mu\text{m/s}$ for the $65 \mu\text{m}$ size range using a stepper actuator (Physik Instrumente M-230S) controlled via a computer interface. Load data were acquired from a 10 g load cell (Transducer Techniques GSO-10) via a DAQ card (PCI-MIO-16E-4) and associated software from National Instruments. Images of the capsule during the compression cycle were captured through a stereo microscope (Nikon SMZ-2T)

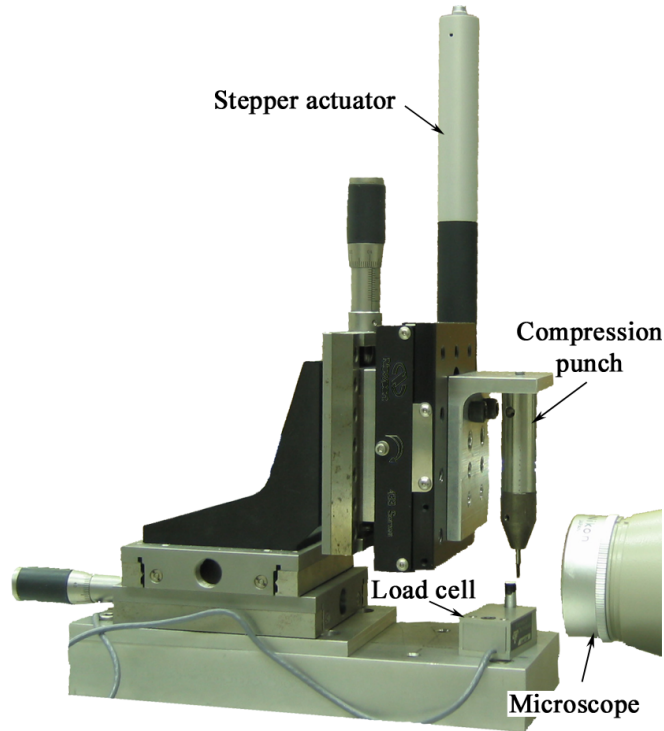


FIG. 3: Photograph of experimental setup.

by a monochrome CCD Camera (Qimaging Retiga). The entire system was mounted on a vibration isolation table.

For a dry microcapsule test, capsules were drawn into a pipette, which enabled release of a single capsule onto the compression platen. An image of the microcapsule was taken prior to compression to determine the initial capsule diameter. An initial separation between the capsule and punch allowed the stepper to achieve steady-state velocity after motion was initiated. The test program was started after positioning the punch above the capsule and terminated after burst was observed.

Immersion tests were conducted using a modification of the apparatus presented in Fig. 3. A schematic of the modified compression setup and the immersion test cell are shown in Fig. 4. Capsules tested in the immersion setup were dispersed in a bath of DCPD and allowed to equilibrate for at least 24 hours. Then a single capsule was removed from solution by pipetting and placed into the compression cell. Fluid was added to the cell cavity to ensure that the entire capsule was submerged. Testing then proceeded as described for the dry test.

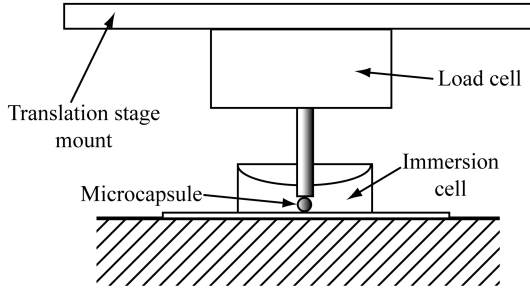


FIG. 4: Schematic of the immersion testing apparatus.

III. COMPRESSION TEST RESULTS

Figure 5 shows representative load–displacement data for an immersed capsule, $213\ \mu\text{m}$ in diameter, tested in compression. Dimensionless displacement is defined by the displacement, δ , divided by the initial capsule diameter, D . Capsules tested while immersed in the encapsulant fluid were imaged by backlighting. In Fig. 5 the images numbered one through four show the capsule during representative sections of the loading sequence. In these images, the solid white line indicates the bottom of the compression platen and the dark region is the compression punch. Image 1 shows the capsule prior to contact with the compression platen. Image 2 is the capsule near the 15% dimensionless displacement point. At approximately this displacement, other researchers have observed a ‘yield’ in the load–displacement response. This yield is characterized by a change in concavity of the load–displacement curve [13]. Image 3 captures the capsule near the ‘burst’ event, which is indicated by the load peak, and image 4 is the capsule after burst.

From Fig. 5 two key observations of compressive capsule shell wall behavior are evident: the capsule shell wall does not buckle during compression and, the capsule remains effectively intact after burst. The absence of buckling indicates that the yield point is due to localized damage, such as microcracking or shear yielding, of the shell wall material. The burst event was also studied with dry compression tests utilizing capsules filled with dyed DCPD. Capsule burst was observed to proceed from the edge of the contact zone where the radius of curvature is the highest. This burst is not a dynamic event, but a leaking of encapsulant fluid from a shell wall failure. The encapsulant leakage proceeds quickly, coating the surface of the capsule in a few seconds.

Figure 6 is a comparison of compression results for dry and immersed capsules of similar diameters. The load–displacement responses of these tests are quite similar in the elastic

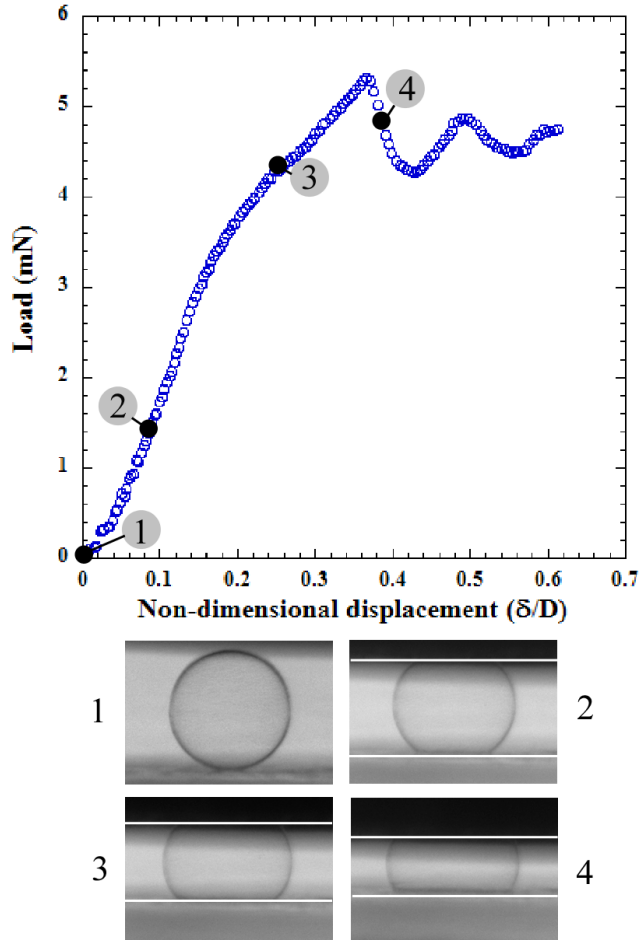


FIG. 5: Images of a $213 \mu\text{m}$ diameter capsule during an immersed compression test.

region of the test (dimensionless displacement less than 15%). At about 20% dimensionless displacement, however, the capsule response changes. The dry capsules generally sustain more load prior to burst than the immersed capsules.

Capsules from batches with average diameters of $187 \pm 15 \mu\text{m}$ and $65 \pm 7 \mu\text{m}$ were tested in dry compression to determine the effect of capsule size. Figure 7 shows representative load–displacement responses for each of the capsule diameters. Two size effects can be noted: smaller capsules are less stiff as a system, in the sense that they sustain less load for a given dimensionless displacement, and the maximum load at burst is highly diameter dependent. The dependence of burst strength on capsule diameter has been observed previously [13]. Table I shows the average burst force and burst strength of the microcapsule types tested. The burst strength is calculated as the burst force normalized by capsule cross-sectional area. The strength data indicate that during dry compression, smaller capsules are harder

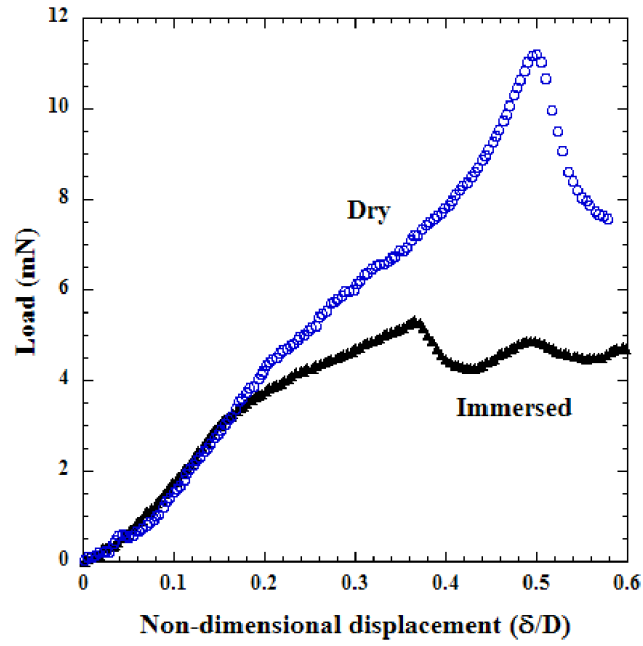


FIG. 6: Comparison of dry and immersed tests on capsules of similar diameter ($222 \mu\text{m}$).

to burst than their larger counterparts.

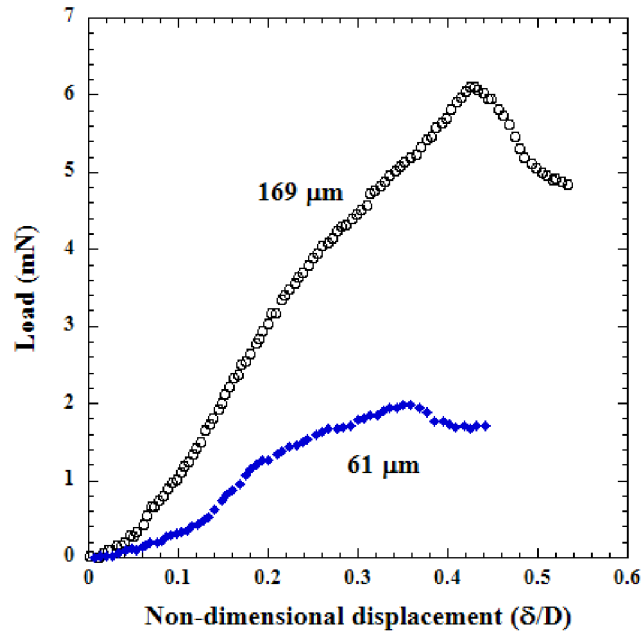


FIG. 7: Comparison of load-displacement responses for a $169 \mu\text{m}$ diameter microcapsule and a $61 \mu\text{m}$ diameter microcapsule tested in dry compression.

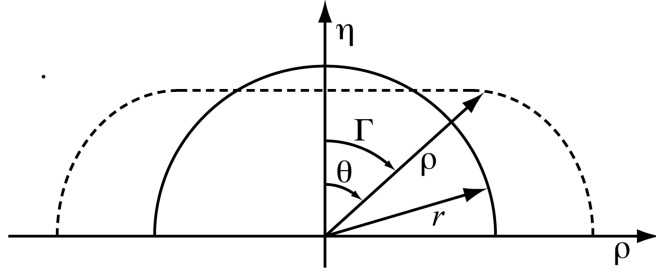


FIG. 8: Schematic of the microcapsule compression problem. The solid line is the uncompressed microcapsule and the dashed line is the compressed microcapsule, after the figure in [12].

IV. COMPARISON WITH THEORY

A. Model Development

The elastic modulus of the capsule shell wall material is extracted through comparison with an analytical membrane theory model. The model is based on the theory initially developed by Feng and Yang [16] for an inflated spherical membrane and then later extended to fluid-filled shells by Lardner and Pujara [17]. The current work follows the analysis for fluid-filled shells as presented in [18] that includes a linear elastic constitutive relationship for the shell wall material. A summary of the model is presented below along with comparisons with the current compression data.

The compression of a microcapsule is shown schematically in Fig. 8, where the solid line is the uncompressed profile and the dashed line represents the compressed geometry. This geometrical arrangement generates two systems of ODEs, one for each distinct region of the compressed capsule. The contact region, the flat portion of the dashed profile, is constrained in one dimension by the compression punch. The non-contact region, which comprises the rest of the capsule shell, is unconstrained and can deform freely.

The system of ODEs for the contact region are

$$\lambda_1' = -\frac{\lambda_1}{\lambda_2 \sin \psi} \frac{f_3}{f_1} - \frac{\lambda_1 - \lambda_2 \cos \psi}{\sin \psi} \frac{f_2}{f_1}, \quad (1)$$

$$\lambda_2' = \frac{\lambda_1 - \lambda_2 \cos \psi}{\sin \psi}, \quad (2)$$

where ψ is the angular coordinate of the compressed capsule, λ_1 and λ_2 are the principal

stretch ratios, and f_1 , f_2 , and f_3 are

$$f_1 = \frac{\partial T_1}{\partial \lambda_1}, \quad (3)$$

$$f_2 = \frac{\partial T_2}{\partial \lambda_2}, \quad (4)$$

$$f_3 = T_1 - T_2, \quad (5)$$

where T_1 and T_2 are the membrane tensions.

The ODEs for the non-contact region are

$$\lambda_1' = \frac{\delta \cos \psi - \omega \sin \psi}{\sin^2 \psi} \frac{f_2}{f_1} - \frac{\omega f_3}{\delta f_1}, \quad (6)$$

$$\delta' = \omega \quad (7)$$

$$\omega' = \frac{\lambda_1' \omega}{\lambda_1} + \frac{\lambda_1^2 - \omega^2}{\delta} \frac{T_2}{T_1} - \frac{\lambda_1 (\lambda_1^2 - \omega^2)^{1/2} P r_0}{T_1}, \quad (8)$$

where

$$\delta = \lambda_2 \sin \psi. \quad (9)$$

A linear elastic constitutive relationship was assumed for the current microcapsule systems and was derived following [18]. The linear-elastic strain energy formula is

$$W = \frac{E h_0}{2(1 + \nu^2)} \left\{ (\lambda_1 - 1)^2 + (\lambda_2 - 1)^2 + 2\nu(\lambda_1 - 1)(\lambda_2 - 1) \right\} \quad (10)$$

from [19]. The wall tensions T_i are related to the strain energy by

$$T_i = \frac{1}{\lambda_1 \lambda_2} \frac{\partial W}{\partial \lambda_i} (\lambda_i)^2. \quad (11)$$

Equation (10) with Eqn. (11) yields

$$T_1 = \frac{E h_0}{(1 - \nu^2)} \frac{\lambda_1}{\lambda_2} \{ (\lambda_1 - 1) + \nu(\lambda_2 - 1) \}, \quad (12)$$

$$T_2 = \frac{E h_0}{(1 - \nu^2)} \frac{\lambda_2}{\lambda_1} \{ (\lambda_2 - 1) + \nu(\lambda_1 - 1) \} \quad (13)$$

for the shell wall tensions. The boundary conditions for this problem are

$$\begin{aligned} \psi = 0 : \quad & \lambda_1 = \lambda_2 = \lambda_0, \\ \psi = \Gamma : \quad & \lambda_{1,\text{contact}} = \lambda_{1,\text{non-contact}}, \\ \psi = \Gamma : \quad & \lambda_{2,\text{contact}} = \lambda_{2,\text{non-contact}}, \\ \psi = \Gamma : \quad & \eta' = 0, \\ \psi = \frac{\pi}{2} : \quad & \delta' = 0. \end{aligned} \quad (14)$$

The above system of ODEs were solved using the numerical scheme outlined in [16]. Numerical solutions were performed with a Runge–Kutta solver provided by Matlab (The MathWorks Inc.). Input to this program consisted of the measured capsule diameter and the average wall thickness. The Poisson’s ratio ν was assumed to be 1/3, a value consistent with other formaldehyde-based polymers. Since the problem is time independent, the load–displacement plots were generated by solving the equilibrium problem of the capsule for a given contact radius and determining the corresponding cross-head displacement.

A sensitivity study was undertaken to investigate the influence of Poisson’s ratio variance on the calculated load–displacement response. Figure 9(a) shows the non-dimensional force ($F = P/Eh_0r_0$)–dimensionless displacement response for different values of the Poisson’s ratio. The model exhibits little sensitivity to Poisson’s ratio, and the results shown in Fig. 9(a) are independent of capsule diameter or shell wall thickness. The sensitivity to shell wall thickness variation was also investigated numerically. Figure 9(b) contains predicted load–displacement curves for a hypothetical capsule with a diameter of 180 μm and a shell wall modulus of 3.6 GPa. The family of curves are the calculated load–displacement responses if the shell wall is assumed to be the average thickness, the upper thickness limit, and the lower thickness limit measured by SEM on a microcapsule batch. These numerical studies indicate that variation in the shell wall thickness alters the predicted modulus value by the same percentage as the thickness variation. That is, if the shell wall is assumed to be 20% thicker than the actual value, the modulus will be under-predicted by 20%. Additionally, the encapsulated volume is assumed constant, as in previous studies [12, 17]. The constant volume assumption disallows fluid diffusion through the shell wall.

B. Property Extraction

Representative model fits for a 169 μm capsule and a 61 μm capsule tested in dry compression are shown in Fig. 10(a). The model was fit to the experimental data using Young’s modulus as the single adjustable parameter. Modulus values obtained from both dry and immersed compression tests are summarized in Table I. The model fits show good agreement until the capsules reach displacement values near 15% (yield point) and then the model deviates significantly from the experimental data. The model predicts a maximum of 3 to 4 percent strain in the capsule shell wall at this point. Strains of this magnitude are sufficient to initiate damage in other thermosetting polymers and, as mentioned previously, the

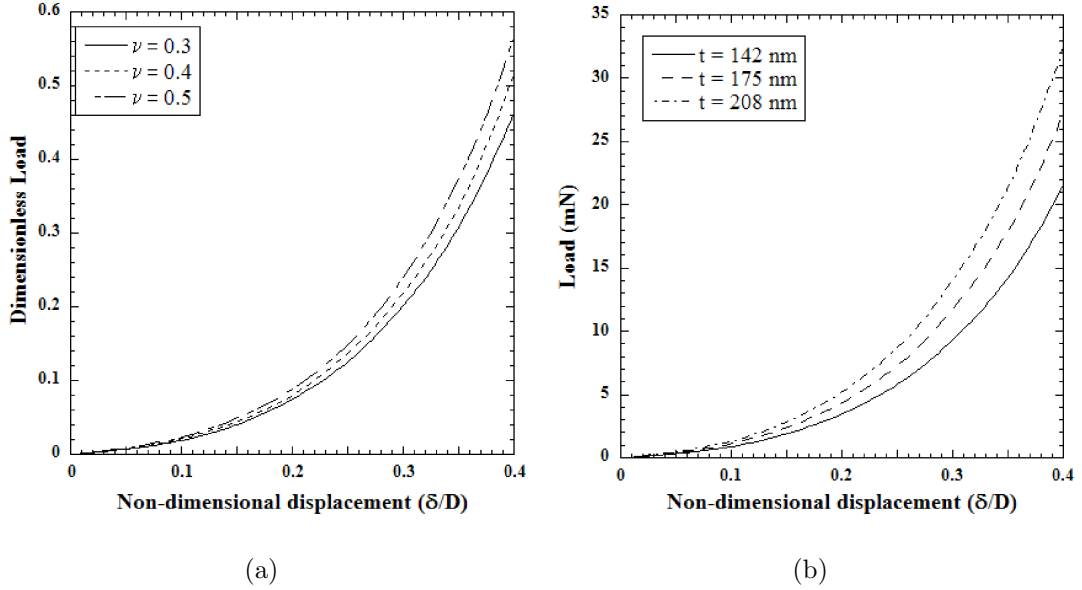
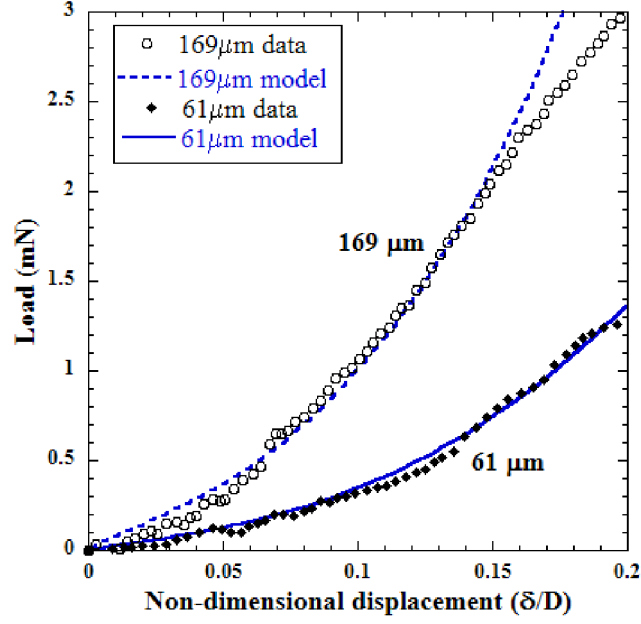


FIG. 9: load–displacement plots from model parameter studies on the (a) effect of Poisson’s ratio ($t = 175$ nm) (b) effect of shell wall thickness variation for a hypothetical $180 \mu\text{m}$ capsule with a shell wall modulus of 3.6 GPa.

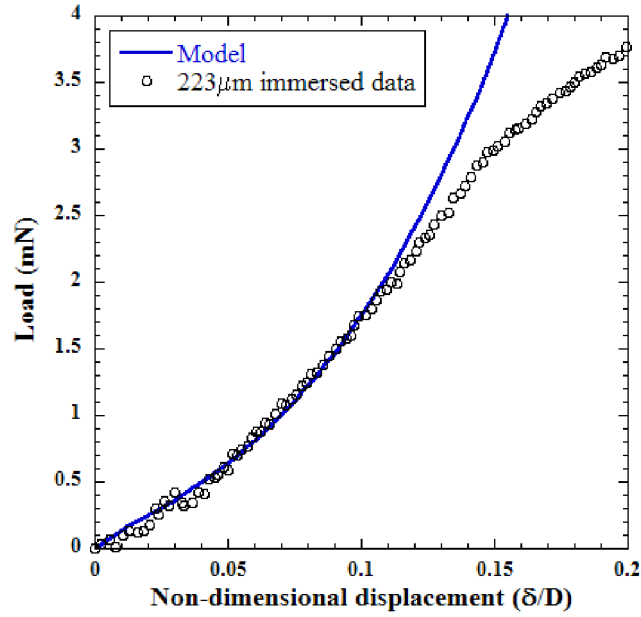
TABLE I: Average Young’s modulus and burst behavior of tested microcapsules.

Average diameter ($\mu\text{m} \pm \text{Std. dev.}$)	t/D	E (GPa $\pm \text{Std. dev.}$)	Average burst force (mN $\pm \text{Std. dev.}$)	Normalized burst strength (MPa $\pm \text{Std. dev.}$)
187 ± 15 , dry	0.001	3.6 ± 0.4	6.5 ± 1.6	0.24 ± 0.04
213 ± 12 , immersed	0.001	3.9 ± 0.7	4.9 ± 0.5	0.14 ± 0.02
65 ± 7 , dry	0.004	3.7 ± 0.5	2.7 ± 0.7	0.8 ± 0.3

yield point may indicate the onset of damage in the shell wall. Dye tests have indicated that there is significant leakage of encapsulant fluid only near 45% deformation, but some diffusion of encapsulant fluid may be occurring. This diffusion would have an effect on the load–displacement behavior of the capsules and is not accounted for in the model. A representative model fit for the compression of a $223 \mu\text{m}$ immersed capsule, Fig. 10(b), shows similar behavior to the dry tests. In this case, the model again deviates just prior to 15% deformation.



(a) Model comparisons for dry compression tests.



(b) Model comparison for an immersed test.

FIG. 10: Comparisons of experimental load–displacement data with the fluid-filled model for dry and immersed UF capsules.

V. CONCLUSIONS

The shell wall elastic modulus of poly(urea-formaldehyde) shelled microcapsules was successfully extracted from single capsule compression testing by comparison with a membrane theory model. Young’s modulus was found to have an average value of 3.7 ± 0.2 GPa for all

capsule testing conditions and was independent of capsule diameter. However, capsule burst behavior was found to be highly diameter dependent. Capsules of smaller diameters, while bursting at lower loads, have a higher specific strength.

Acknowledgments

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